

# RNA Tertiary Structure Mediation by Adenosine Platforms

Jamie H. **Cate**, Anne R. Gooding, Elaine Podell, Kaihong Zhou, Barbara L. Golden, Alexander A. Szewczak, Craig E. Kundrot, Thomas R. Cech, Jennifer A. Doudna \*

The crystal structure of a group I intron domain reveals an unexpected motif that mediates both intra- and intermolecular interactions. At three separate locations in the 160-nucleotide domain, adjacent adenosines in the sequence lie side-by-side and form a pseudo-base pair within a helix. This adenosine platform opens the minor groove for base stacking or base pairing with nucleotides from a noncontiguous RNA strand. The platform motif has a distinctive chemical modification signature that may enable its detection in other structured RNAs. The ability of this motif to facilitate higher order folding provides one explanation for the abundance of adenosine residues in internal loops of many RNAs.

J. H. **Cate**, K. Zhou, J. A. Doudna, Department of Molecular Biophysics and Biochemistry, Yale University, New Haven, CT 06520.

A. R. Gooding, E. Podell, B. L. Golden, A. A. Szewczak, T. R. Cech, Department of Chemistry and Biochemistry and Howard Hughes Medical Institute, University of Colorado, Boulder, CO 80309.

C. E. Kundrot, Department of Chemistry and Biochemistry, University of Colorado, Boulder, CO 80309.

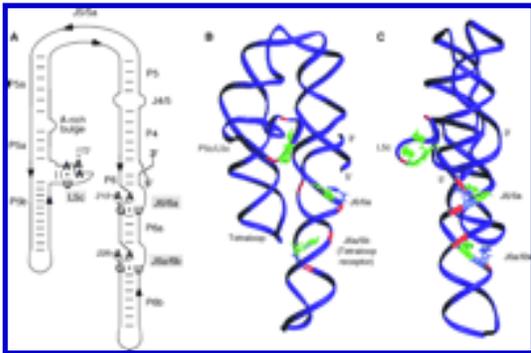
\* To whom correspondence should be addressed.

- ▶ [Abstract of this Article](#)
- ▶ Similar articles found in: [SCIENCE Online](#) [PubMed](#)
- ▶ [PubMed Citation](#)
- ▶ This Article has been cited by: [other online articles](#)
- ▶ Search Medline for articles by: [Cate, J. H.](#) || [Doudna, J. A.](#)
- ▶ Alert me when: [new articles cite this article](#)
- ▶ [Download to Citation Manager](#)

- ▶ Collections under which this article appears: [Molecular Biology](#)

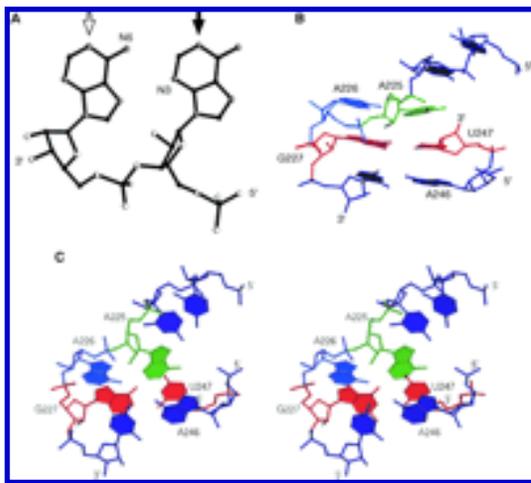
Ribozymes and large RNA components of spliceosomes and ribosomes fold into complex three-dimensional architectures. To form these biologically active structures, helical regions must pack together specifically. Comparative sequence analysis (1, 2), biochemical experiments (2, 3, 4, 5), and modeling based on intermolecular contacts in crystals of small RNAs (6) have identified some elements responsible for long-range tertiary interactions in large RNAs, but their molecular details are largely unknown. The crystal structure of the 160-nucleotide P4-P6 domain of the *Tetrahymena thermophila* self-splicing intron (7) has revealed several new types of long-range contacts, including three examples of the adenosine platform motif described below.

The secondary structure of the P4-P6 domain, like that of many other large RNAs, contains base-paired regions interspersed with internal loops (Fig. 1A). As in other RNAs, many of the loops contain a high proportion of adenosines (8). At three different sites in the crystal structure of the domain, two such adenosines assume a side-by-side configuration which we call an adenosine platform, or A-A platform (Fig. 1, B and C). The A-A platform motif consists of adjacent adenosines in a helical strand that form a pseudo base pair within the helix (Fig. 2A). The 3' A continues the stacking of the A-form helix below it, and the 5' A stacks on the opposite strand of the helix (Fig. 2, B and C). Although the two adenosines may share one hydrogen bond (9, 10), the driving force for the platform appears to be base stacking. A non-Watson-Crick base pair below each platform (G·U in two cases, a non-Watson-Crick A·U in the third) shifts the base positions to increase stacking with the two adenosines of the platform. In addition, the ribose of the 5' A has a noncanonical pucker (11), facilitating a kink in the phosphate backbone that broadens the minor groove in the dimension parallel to the helix axis. The overall geometries of the platforms are similar (rmsd, 0.5 to 1.2 Å), although we observe differences in the coordination of water and metal ions in the major grooves of the motifs (12).



**Fig. 1.** (A) Schematic secondary structure of the P4-P6 domain, with adenosine platforms in boldface. The noncanonical base pair below each platform is also shown. (B) Location of adenosine platforms in the crystal structure. The structure is shown in the same orientation as that of (A); adenosines of the platforms are highlighted in blue and green, while the backbone positions of the wobble base pairs below the platforms are

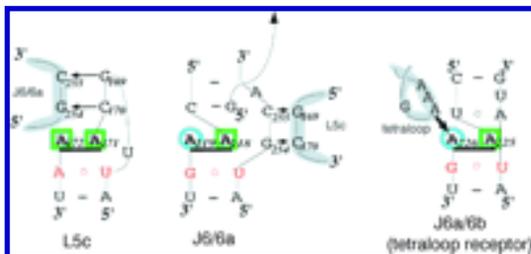
indicated in red. Blue adenosines are accessible to dimethyl sulfate in the presence and absence of the docking partner associated with the platform; green adenosines are protected from dimethyl sulfate modification when the docking partner is present (see text and Fig. 3 for details). (C) Another view of the crystal structure, rotated 90° about the long axis of the molecule. The conserved core region is to the right in (A) and (B) and the core faces the reader in (C). This figure was prepared by means of the molecular graphics program RIBBONS (24). [\[View Larger Version of this Image \(31K GIF file\)\]](#)



**Fig. 2.** (A) View of an adenosine platform looking down the helix axis. N3 of the 5<sup>′</sup> A and N6 of the 3<sup>′</sup> A are within hydrogen bonding distance (2.8 to 3.4 Å). Closed arrow, N1 position of the 5<sup>′</sup> A, which is protected from dimethyl sulfate when the long-range contact is formed; open arrow, N1 position of the 3<sup>′</sup> A which shows variable dimethyl sulfate protection (see 18). (B) The adenosine platform in the tetraloop receptor; color scheme as in Fig. 1. (C) Stereo view from underneath the platform, looking up the helix axis. Figure prepared with RIBBONS (24).

[\[View Larger Version of this Image \(31K GIF file\)\]](#)

Each adenosine platform mediates a long-range contact in the RNA (Fig. 3). One of the contacts is intramolecular, and the other two occur between adjacent molecules in the crystal lattice. The intramolecular contact contributes to a key component of the domain tertiary structure. Contained within the tetraloop receptor motif found in group I introns and other RNAs (2), the platform in J6a/6b opens the minor groove to enable the GAAA tetraloop of the same molecule to dock above the platform. This interaction helps hold together the two helical halves of the molecule (7). The other two platforms, in L5c and J6/6a, are involved in packing the two molecules into the asymmetric unit of the crystal, perhaps mimicking contacts between different domains in the intact intron. In the crystal, two G·C base pairs connect J6/6a of molecule A to L5c of molecule B and vice versa (Fig. 3). The platform in J6/6a stacks in the helix as if it were a base pair. This allows nucleotides in the strand opposite the platform to flip away from the helix and base pair with nucleotides from L5c in the neighboring molecule. The helix and stacking in J6/6a continue above the platform with a C·G base pair formed between the strand of the platform and a G at the 5<sup>′</sup> end of the molecule (Fig. 3). The platform in L5c reciprocates the base-pairing interaction with J6/6a. In this case, however, the platform caps a helix, allowing the intermolecular base pairs to stack on top; the helix is effectively extended two base pairs by the long-range contact.



**Fig. 3.** The different kinds of long-range interactions that occur near the adenosine platforms. At left and in the center, reciprocal interactions occur between L5c and J6/6a in the two molecules in the asymmetric unit of the crystal; at right, the tetraloop docks above the platform in the tetraloop receptor. The docking partner for each platform is shaded. Blue adenosines (circles) are accessible to dimethyl sulfate in the presence and absence of the docking partner associated with the platform. Green adenosines (squares) are protected from dimethyl sulfate modification when the docking partner is present. The noncanonical base pair below each platform is highlighted in red.

The docking partner for each platform is shaded. Blue adenosines (circles) are accessible to dimethyl sulfate in the presence and absence of the docking partner associated with the platform. Green adenosines (squares) are protected from dimethyl sulfate modification when the docking partner is present. The noncanonical base pair below each platform is highlighted in red.

[\[View Larger Version of this Image \(18K GIF file\)\]](#)

Mutational analysis supports the functional role of two of the A-A platform motifs. Activity of P4-P6 variants can be assessed in vitro in a three-component ribozyme in which the P4-P6 domain assembles with the rest of the intron via tertiary interactions (13). Mutation of the A-A platform in J6a/6b (the tetraloop receptor), which prevents its interaction with the tetraloop and thereby destabilizes the P4-P6 domain (5), reduced activity 70-fold (14). Removal of the A-A platform motif in J6/6a has no discernible effect on the overall structure of the P4-P6 domain (5), yet ribozyme activity dropped 25-fold (14). These results suggest that the A-A platform in J6/6a contributes to interdomain association, while that in J6a/6b contributes to P4-P6 structure and perhaps additionally to its association with other regions of the ribozyme. While the three-component functional assay was performed in 80 mM MgCl<sub>2</sub>, a condition that suppresses many ribozyme mutations (15, 16), the two A-A platform mutants still showed significantly reduced activity.

A consistent pattern of dimethyl sulfate modification of the RNA is observed at each A-A platform. When the intact *Tetrahymena* intron is probed with dimethyl sulfate, the 5<sup>ʹ</sup> adenosine of each platform is protected from methylation at its N1 position (17). In the P4-P6 domain by itself, the 5<sup>ʹ</sup> As of the platforms in J6/6a and L5c are susceptible to methylation (17). The 5<sup>ʹ</sup> A of the platform in J6a/6b (the tetraloop receptor) also becomes susceptible to N1-methylation by dimethyl sulfate when the tetraloop-receptor interaction is disrupted by mutation (17, 18). Thus in each case, the 5<sup>ʹ</sup> A is protected from dimethyl sulfate only when the associated long-range contact is present. The unusual geometry of the A-A platforms seen in the crystal structure explains these results. Although there are no apparent hydrogen bonds to the Watson-Crick face of the 5<sup>ʹ</sup> A in the platform, the base is buried in the helix and the N1 position is inaccessible (Fig. 2A) (19). The fact that the 5<sup>ʹ</sup> A in the platform becomes accessible to dimethyl sulfate when its docking partner is removed suggests that the platform structure is dynamic or undergoes a conformational change in the absence of the long-range interaction. Formation of a stable structure only upon binding of a ligand is well preceded from NMR structures of small RNA motifs (20). Although the methylation data clearly implicate the J6/6a and L5c platforms as having binding partners in a portion of the intron outside the P4-P6 domain, these tertiary interactions have not yet been located.

A comparison of potential adenosine platforms in similar group I introns [subclasses IC1 and IC3 (2, 21)] reveals that, whereas A-A platforms are most common (83/93 potential platforms surveyed), variation occurs in both positions. The stacking interactions that stabilize the platform would be expected to favor purines at the platform positions, yet only one potential G-G platform occurs. Pyrimidines occur at one or both positions in eight cases. Whether these variations are functionally significant is unclear in that no activity information is available for the introns in question (22).

The adenosine platform motif provides one explanation for the abundance of adenosines in internal loops of many large RNAs. Since stacking may be the primary energetic component of the platform, the adenine base is perhaps particularly well suited because it stacks efficiently while minimizing the potential for steric clash. Whether adenosine platforms are a common structural stepping stone to higher order RNA folding is uncertain, but the tetraloop receptor, which contains an adenosine platform, is

widespread in the group I and group II families of introns (2). The invariant A-A dinucleotide at the A site in the ribosome (nucleotides 1492 and 1493 in *Escherichia coli* 16S ribosomal RNA) exhibits a dimethyl sulfate protection pattern characteristic of the platform motif when probed in the presence and absence of tRNA (23). Use of an adenosine platform to mediate tRNA binding at a fundamental step of translation would suggest that adenosine platforms arose early in evolution as an efficient mechanism for building complex, functional RNA architectures.

## REFERENCES AND NOTES

1. F. Michel and E. Westhof, *J. Mol. Biol.* **216**, 585 (1990) [[Medline](#)].
2. M. Costa and F. Michel, *EMBO J.* **14**, 1276 (1995) [[Medline](#)].
3. L. Jaeger, E. Westhof, F. Michel, *J. Mol. Biol.* **221**, 1153 (1991) [[Medline](#)].
4. M. E. Harris *et al.*, *EMBO J.* **13**, 3953 (1994) [[Medline](#)].
5. F. L. Murphy and T. R. Cech, *J. Mol. Biol.* **236**, 49 (1994) [[Medline](#)].
6. H. Pley, K. Flaherty, D. McKay, *Nature* **372**, 111 (1994) [[Medline](#)].
7. J. H. Cate *et al.*, *Science* **273**, 1678 (1996) .
8. C. R. Woese, S. Winker, R. R. Gutell, *Proc. Natl. Acad. Sci. U.S.A.* **87**, 8467 (1990) [[Medline](#)].
9. The N3 of the 5' adenosine is 2.8 to 3.4 Å from the N6 of the 3' adenosine in the platform; coordinate error in the current model is approximately 0.4 Å, determined according to P. V. Luzzati [*Acta Crystallogr.* **5**, 802 (1955)].
10. K. Shetty and S. A. Strobel, unpublished data.
11. In the two platforms in internal loops (J6/6a and J6a/6b), the 5'-A pucker is O4'-endo, C1'-exo; in L5c the 5'-A of the platform has C3'-exo sugar pucker. Canonical A-form helices have C3'-endo ribose puckers; *W. Saenger, Principles of Nucleic Acid Structure* (Springer-Verlag, New York, 1984), pp. 55-68.
12. Comparisons between platforms in the internal loops (J6/6a and J6a/6b) involved superposition of all atoms in the A-A and the G·U wobble below. Comparisons between the L5c platforms in the two molecules in the asymmetric unit included the analogous four residues; comparisons between the L5c and internal bulge platforms included only the adenosines of the platform and the 3' purine in each case. In both intrahelical platforms (J6/6a and J6a/6b), strong density in the  $F_0 - F_c$  electron density map is consistent with a metal ion or water molecule in the major groove coordinated to the ribose and phosphate of the 5' A and 3' A of the platform, respectively, and to N7 and O6 of the G in the G·U wobble below the motif. In the L5c loop platform, however, there is density for a putative magnesium ion coordinated to phosphates of the 3' A of the platform and the adjacent A of the A·U noncanonical pair. Both molecules in the asymmetric unit reflect these differences between the platforms.
13. J. A. Doudna and T. R. Cech, *RNA* **1**, 36 (1995) [[Medline](#)].
14. The J6a/6b paired mutant has U224-A225-A226 converted to an A-U dinucleotide, causing it to base pair with the A-U across the loop. It showed a cleavage rate of  $0.006 \pm 0.003 \text{ min}^{-1}$  (mean  $\pm$  range of four experiments) with 100 nM of each domain, a concentration that is nearly

saturating for the wild-type components ( $K_m = 31$  nM for P4-P6 and 4 nM for P3-P9). The J6/6a paired mutant has C217-A218-A219 converted to the sequence U-G-C, causing it to base pair with the G-C-A across the loop. It showed a cleavage rate of  $0.0166 \pm 0.0004$  min<sup>-1</sup> under the conditions described above. Wild-type P4-P6 gave a rate of  $0.43 \pm 0.04$  min<sup>-1</sup> in side-by-side experiments. Preliminary RNA splicing analysis of a precursor RNA with a two-base change in the third adenosine platform, A171-A172 to U-U, showed a twofold reduction in activity at low magnesium ion concentration (5 mM) and even less of an effect at higher magnesium concentrations. One example each of potential C-U and U-C platforms occurs in the tetraloop receptor motif of group I introns in subclasses IC1 and IC3.

15. S. Couture *et al.*, *J. Mol. Biol.* **215**, 345 (1990) [[Medline](#)].
16. B. Laggerbauer, F. L. Murphy, T. R. Cech, *EMBO J.* **13**, 2669 (1994) [[Medline](#)].
17. F. L. Murphy and T. R. Cech, *Biochemistry* **32**, 5291 (1993) [[Medline](#)].
18. In the internal-loop A platforms, the 3<sup>rd</sup> A is always susceptible to methylation. In the L5c platform, the 3<sup>rd</sup> A is protected only in the intact intron.
19. The solvent accessible surface of the molecule was determined with a probe of radius 2.0 for adenosine modification Å, the "effective" radius of dimethyl sulfate determined by the method of S. R. Holbrook and S.-H. Kim [*Biopolymers* **22**, 1145 (1983)].
20. J. D. Puglisi, R. Tan, B. J. Calnan, A. D. Frankel, J. R. Williamson, *Science* **257**, 76 (1992) [[Medline](#)]; F. Aboul-ela, J. Karn and G. Varani, *J. Mol. Biol.* **253**, 313 (1995) ; Y. Yang, M. Kochoyan, P. Burgstaller, E. Westhof, M. Famulok, *Science* **272**, 1343 (1996) [[Medline](#)]; F. Jiang, R. A. Kumar, R. A. Jones, D. J. Patel, *Nature* **382**, 171 (1996) ; T. Dieckmann, E. Suzuki, G. K. Nakamura, J. Feigon, *RNA*, in press.
21. S. H. Damberger and R. R. Gutell, *Nucleic Acids Res.* **22**, 3508 (1994) [[Medline](#)].
22. Mutation of the J6a/6b platform in P4-P6 from A-A to G-A produced a P4-P6 derivative that appeared to fold correctly in solution (5), but the RNA was not tested for activity.
23. D. Moazed and H. F. Noller, *Cell* **47**, 985 (1986) [[Medline](#)].
24. M. Carson, *J. Appl. Cryst.* **24**, 958 (1991).
25. We thank K. Blount, D. Sheehan, and A. Zaug for functional assays; F. Michel, P. Moore, and A. M. Pyle for helpful discussions; and A. Ferré-d'Amaré, S. Strobel, and T. Griffin for review of the manuscript. This work was funded by the Lucille P. Markey Charitable Trust, the Donaghue Medical Research Foundation and NIH grant GM22778-21 (J.A.D.), NIH training grant 5T32GM08283-07 (J.H.C.), American Cancer Society postdoctoral fellowship (B.L.G.), NSF grant MCB-9221307 (C.E.K.), Howard Hughes Medical Institute (T.R.C.), and the Keck Foundation (C.E.K. and T.R.C.); T.R.C. is a Howard Hughes Medical Institute Investigator; J.A.D. is a Lucille P. Markey Scholar in Biomedical Science, a Young Investigator of the Donaghue Medical Research Foundation, a Searle Scholar, and a Beckman Young Investigator.

15 July 1996; accepted 16 August 1996

- ▶ [Abstract of this Article](#)
- ▶ Similar articles found in:  
[SCIENCE Online](#)  
[PubMed](#)
- ▶ [PubMed Citation](#)
- ▶ This Article has been cited by:
- ▶ Search Medline for articles by:  
[Cate, J. H.](#) || [Doudna, J. A.](#)
- ▶ Alert me when:  
[new articles cite this article](#)
- ▶ [Download to Citation Manager](#)

- ▶ Collections under which this article appears:  
[Molecular Biology](#)

## This article has been cited by other articles:

- [Cate, J. H.](#), Gooding, A. R., Podell, E., Zhou, K., Golden, B. L., Kundrot, C. E., Cech, T. R., Doudna, J. A. (1996). Crystal Structure of a Group I Ribozyme Domain: Principles of RNA Packing. *Science* 273: 1678-1685 [[Abstract](#)] [[Full Text](#)]
- Thirumalai, D. (1998). Native secondary structure formation in RNA may be a slave to tertiary folding. *Proc. Natl. Acad. Sci. U. S. A.* 95: 11506-11508 [[Full Text](#)]
- Conn, G. L., Draper, D. E., Lattman, E. E., Gittis, A. G. (1999). Crystal Structure of a Conserved Ribosomal Protein-RNA Complex. *Science* 284: 1171-1174 [[Abstract](#)] [[Full Text](#)]
- Narlikar, G. J., Herschlag, D. (1997). MECHANISTIC ASPECTS OF ENZYMATIC CATALYSIS: Lessons from Comparison of RNA and Protein Enzymes. *Annu. Rev. Biochem.* 66: 19-59 [[Abstract](#)] [[Full Text](#)]
- Wedekind, J. E., McKay, D. B. (1998). CRYSTALLOGRAPHIC STRUCTURES OF THE HAMMERHEAD RIBOZYME: Relationship to Ribozyme Folding and Catalysis. *Annu. Rev. Biophys. Biomol. Struct.* 27: 475-502 [[Abstract](#)] [[Full Text](#)]
- Ferré-D'Amaré, A. R., Doudna, J. A. (1999). RNA FOLDS: Insights from Recent Crystal Structures. *Annu. Rev. Biophys. Biomol. Struct.* 28: 57-73 [[Abstract](#)] [[Full Text](#)]
- Moore, P. B. (1999). STRUCTURAL MOTIFS IN RNA. *Annu. Rev. Biochem.* 68: 287-300 [[Abstract](#)] [[Full Text](#)]
- Golden, B. L., Gooding, A. R., Podell, E. R., Cech, T. R. (1998). A Preorganized Active Site in

- the Crystal Structure of the Tetrahymena Ribozyme. *Science* 282: 259-264 [\[Abstract\]](#) [\[Full Text\]](#)
- Babel, T B, Warnke, P C, Ostertag, C B (2001). Immediate and long term outcome after infrathalamic and thalamic lesioning for intractable Tourette's syndrome. *J. Neurol. Neurosurg. Psychiatry* 70: 666-671 [\[Abstract\]](#) [\[Full Text\]](#)
  - Zhang, Y., Leibowitz, M. J. (2001). Folding of the group I intron ribozyme from the 26S rRNA gene of *Candida albicans*. *Nucleic Acids Res* 29: 2644-2653 [\[Abstract\]](#) [\[Full Text\]](#)
  - Boudvillain, M., Pyle, A. M. (1998). Defining functional groups, core structural features and inter-domain tertiary contacts essential for group II intron self-splicing: a NAIM analysis. *EMBO J.* 17: 7091-7104 [\[Abstract\]](#) [\[Full Text\]](#)
  - Jaeger, L., Wright, M. C., Joyce, G. F. (1999). A complex ligase ribozyme evolved in vitro from a group I ribozyme domain. *Proc. Natl. Acad. Sci. U. S. A.* 96: 14712-14717 [\[Abstract\]](#) [\[Full Text\]](#)
  - Esteban, J. A., Walter, N. G., Kotzorek, G., Heckman, J. E., Burke, J. M. (1998). Structural basis for heterogeneous kinetics: Reengineering the hairpin ribozyme. *Proc. Natl. Acad. Sci. U. S. A.* 95: 6091-6096 [\[Abstract\]](#) [\[Full Text\]](#)
  - Wu, M., Tinoco, I. Jr. (1998). RNA folding causes secondary structure rearrangement. *Proc. Natl. Acad. Sci. U. S. A.* 95: 11555-11560 [\[Abstract\]](#) [\[Full Text\]](#)
  - Jestin, J.-L., Dème, E., Jacquier, A. (1997). Identification of structural elements critical for inter-domain interactions in a group II self-splicing intron. *EMBO J.* 16: 2945-2954 [\[Abstract\]](#) [\[Full Text\]](#)
  - Costa, M., Michel, F. (1997). Rules for RNA recognition of GNRA tetraloops deduced by in vitro selection: comparison with in vivo evolution. *EMBO J.* 16: 3289-3302 [\[Abstract\]](#) [\[Full Text\]](#)
  - Butcher, S. E., Dieckmann, T., Feigon, J. (1997). Solution structure of a GAAA tetraloop receptor RNA. *EMBO J.* 16: 7490-7499 [\[Abstract\]](#) [\[Full Text\]](#)
  - Konforti, B. B., Liu, Q., Pyle, A. M. (1998). A map of the binding site for catalytic domain 5 in the core of a group II intron ribozyme. *EMBO J.* 17: 7105-7117 [\[Abstract\]](#) [\[Full Text\]](#)
  - Atsumi, S., Ikawa, Y., Shiraishi, H., Inoue, T. (2001). Design and development of a catalytic ribonucleoprotein. *EMBO J.* 20: 5453-5460 [\[Abstract\]](#) [\[Full Text\]](#)
  - Burkhardt, C., Zacharias, M. (2001). Modelling ion binding to AA platform motifs in RNA: a continuum solvent study including conformational adaptation. *Nucleic Acids Res* 29: 3910-3918 [\[Abstract\]](#) [\[Full Text\]](#)
  - Barrick, J. E., Takahashi, T. T., Ren, J., Xia, T., Roberts, R. W. (2001). Large libraries reveal diverse solutions to an RNA recognition problem. *Proc. Natl. Acad. Sci. U. S. A.* 98: 12374-12378 [\[Abstract\]](#) [\[Full Text\]](#)

Volume 273, Number 5282, Issue of 20 Sep 1996, pp. 1696-1699.

Copyright © 1996 by The American Association for the Advancement of Science.



 **PAGE TOP**